Representing and Comparing Metabolic Pathways as Petri Nets with MPath2PN and CoMeta

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Abstract

We present two tools, *MPath2PN* and *CoMeta*, which are part of an ongoing project for representing and comparing metabolic pathways as Petri Nets. *MPath2PN* is intended to support an automatic translation of metabolic pathways from the major biological databases into corresponding Petri nets expressed in the input formalisms for Petri net tools. *CoMeta* is devised to compare metabolic pathways of different organisms through their Petri net representation produced by *MPath2PN*. *CoMeta* automatically takes the data from the KEGG database and, in the comparison, it considers both homology of reactions and behavioural aspects of the pathways as expressed by the T-invariants of the underlying Petri nets.

Keywords: Metabolic pathways, Petri nets, translation, comparison

1 Introduction

The metabolism is the chemical system which generates the amino acids, sugars, lipids, nucleic acids and energy necessary to the life of any organism. Metabolic pathways are subsystems dealing with specific functions. Their

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comparison in different species yields interesting information on evolution and it may help in understanding the associated functions. This is important for studying diseases and drugs design and for industrial applications. We present two tools which are part of an ongoing project for the representation and comparison of metabolic pathways as basic Petri nets. Preliminary prototypes of such tools have been presented in [20,21].

A metabolic pathway consists of a network of chemical reactions, catalysed by one or more enzymes, where some molecules (reactants or substrates) are transformed into others (products). Enzymes are not consumed in a reaction, even if they are necessary and used while the reaction takes place. The product of a reaction is the substrate of the next one. Quantitative relations can be represented through a stoichiometric matrix, where rows represent molecular species and columns represent reactions. An element of the matrix, a stoichiometric coefficient n_{ij}, represents the degree to which the i-th chemical species participates in the j-th reaction. The kinetic of a pathway is determined by the rate associated to each reaction. Information on metabolic pathways are collected in several databases. In particular the KEGG PATHWAY database [8] (Kyoto Encyclopedia of Genes and Genomes) contains metabolic, regulatory and genetic pathways for different species whose data are derived by genome sequencing. It integrates genomic, chemical and systemic functional information [25] represented as maps which are linked to additional information on reactions, proteins and genes, possibly stored in other databases. KEGG pathways are coded using KGML (KEGG Markup Language) [7] based on XML. A web service for querying the system from users programs is available. KEGG provides a uniform view of the same pathway in different organisms. There are other important free access repositories on metabolic pathways, e.g., the BioModels Database, whose biological models are coded in SBML (Systems Biology Markup Language) [15], and MetaCyc [10], which is part of BioCyc Database Collection [1].

Petri nets (PNs) [28,22] are a well studied formalism offering many tools for visualisation, simulation and analysis (see *The Petri net World site* [12]). They are well suited for metabolic pathways because of the natural correspondence between PNs and biochemical networks [33,32,23] and indeed they have been largely used for qualitative and quantitative modelling and analysis of metabolic pathways (see e.g. [19] for a survey). In a Petri net places represent molecular species, such as metabolites, proteins or enzymes; transitions correspond to chemical reactions; input places represent the substrate or reactants and output places reaction products. The incidence matrix of the PN is identical to the stoichiometric matrix, the number of tokens in each place indicates the amount of substance associated with that place, the flux modes and the conservation relations for metabolites correspond to specific properties of PNs. In particular minimal (semi-positive) T-invariants correspond to elementary

flux modes [34] of a metabolic pathway, i.e., minimal sets of reactions that can operate at a steady state. Minimal T-invariants form a basis for the set of semi-positive T-invariant (Hilbert basis) which is unique and characteristic of the PN.

The rest of the paper is devoted to the presentation of the tools. In Section 2 we describe *MPath2PN*, a tool which supplies an automatic translation of the data on a metabolic pathway into a PN model. It is written in Java and it is conceived to deal with different databases in input, such as KEGG [8] and the BioModels Database [15], and different PN tools formats in output. In Section 3 we describe *CoMeta*, a tool which automatically takes the data from the KEGG database and it compares metabolic pathways of different organisms relying on their PN representation. The comparison is based on a similarity measure which considers both homology of reactions and functional aspects of the pathways. The latter are captured by the (semi-positive) T-invariants of the PNs which correspond to potential fluxes at steady state. Some conclusions are drawn in Section 4.

The presented tools, *Mpath2PN* and *CoMeta* are freely available at the address http://www.dsi.unive.it/~biolab.

2 The Tool *MPath2PN*

The availability of different sources of data on metabolic pathways and of many simulation and analysis tools for PNs paradoxically is a problem, because of the variety of database formats and of input formats for PNs tools. In the literature, in order to cope with this problem, we find proposals for

- a standard format for metabolic data, such as SBML [15], the format of the BioModels Database [2], or BioPAX [3], and a standard format for PN tools, such as PNML [11];

- unification or integration of different databases such as in [31] or [24], and translations between different data formats, such as in KEGGtranslator [9] or KGML2SBML and KGML2BioPAX [26].

Automatic translators from metabolic pathways to PN models are offered as specific functionalities in some modelling and analysis tools based on PNs. Snoopy [16,27], a tool for the design and animation of PNs to verify technical systems and validate biosystems, provides translations from SBML to basic PNs, continuous PNs and extended stochastic PNs in a Snoopy-specific format based on XML. Cell Illustrator (CI)[4,30], a tool for modelling and simulating complex biological systems, allows the user to import models in SBML and BioPAX and translates them into CSML, the XML-based format of CI models, which are Petri nets extended with continuous and generic processes and quantities.
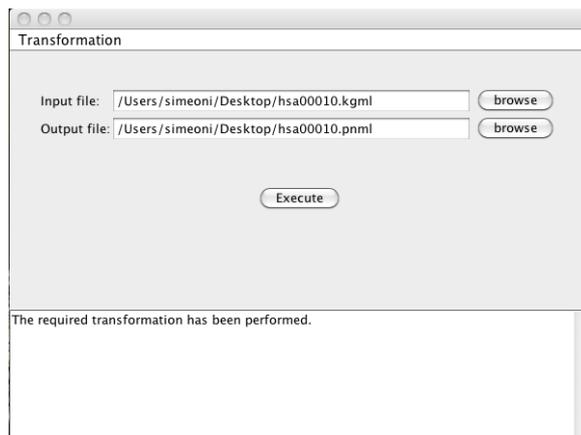
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Fig. 1. The MPATH2PN graphical user interface

We developed the tool *MPath2PN* to support the automatic translation of metabolic pathways into PN models, coping with different input and output formats. *MPath2PN* is written in Java and it has a modular structure which is designed to facilitate the integration of different translations. Since most of the formats for metabolic pathways and for PNs are based on XML, translations are implemented by using XSLT (eXtensible Stylesheet Language Transformation [6]) in the Saxon open source version [14]. Each translation requires the definition of an appropriate style sheet XSL [5] which specifies the translation rules to be applied. For integrating different sources of information the translation process in *MPath2PN* is organised into three phases: pre-treatment, XSLT translation and post-treatment. In the pre- and post-treatment Java classes can be developed for modifying respectively the input and the output files. For example when KEGG data are in input, the KGML file representing a KEGG pathway does not specify the stoichiometric values of reactions. The post-treatment phase can retrieve the missing information from the KEGG web service and add them to the PN model.

The user interface of *MPath2PN*, shown in Figure 1, is very simple. The different translations offered by the tool may be chosen through the "*Transformation*" menu. It is sufficient to specify the input and output file names and press the "*Execute*" button. Information messages on the translation status are displayed in the lower part of the window.

At present *MPath2PN* provides various translations from KEGG metabolic pathways (expressed using KGML) to PNML (Petri Net Markup Language [11]), a standard format for PNs tools. Such translations differ on the adopted modelling choices with respect to the following issues.

- *Representation of ubiquitous substances*: Once assumed to be constant, ubiquitous substances, such as ATP, ADP and H_2O, can be omitted and the resulting model is greatly simplified, but it could become disconnected.

MPath2PN translations either include ubiquitous substances or ignore them.

- *Representation of reactions*: *MPath2PN* translations are either *reaction-based* or *enzyme-based.* In a reaction-based translation, each reaction, possibly catalysed by various enzymes, is represented by a single transition in the PN. In an enzyme-based translation, each reaction is represented by as many transitions as the number of enzymes catalysing it.

- *Representation of boundaries*: A PN corresponding to a metabolic pathway of an organism can be considered either in isolation, focusing on its internal behaviour, or as an open interactive subsystem of the full metabolic network. *MPath2PN* offer the option of producing either *isolated* or *open* PN models. In the open case, compounds which connect the pathway to the rest of the metabolic network are represented as open places, i.e. places where the environment can freely put/remove substances through corresponding input/output transitions. Compounds which are only substrates/only products (the source and the target substances of the pathway) are represented as open places where the environment can freely put/remove substances.

Most of the reactions in a pathway are reversible. In the PN model *MPath2PN* represent a reversible reaction by using two transitions, a forward and a backward one. Such pairs of transitions are recognisable by their identifiers, so that the corresponding cyclic behaviour can be filtered out, if desired, in the model analysis.

A first prototype version of *MPath2PN* was presented in [20], it provided two translations from KEGG to the input language of PIPE2 [13], an open source platform independent tool for creating and analysing PNs. Both translations produce reaction-based, isolated PNs, differing for the treatment of the ubiquitous substances.

3 The tool *CoMeta*

In the literature many proposals for comparing metabolic pathways, or even whole metabolic networks, in different organisms can be found. They are based on two ingredients: a simplified representation of a metabolic pathway, viewed as a set of reactions and/or compounds or as a set of sequences of reactions or as a graph, and a notion of similarity score (or distance measure) between two pathways which depends on the chosen representation. In [21] various approaches to the comparison of metabolic pathways are discussed, pointing out that they are eminently static and ignore the flow of metabolites in the pathways. The tool *CoMeta* (COmparing METAbolic pathways) compares metabolic pathways of different organisms, considering also their dynamics, relying on a PN representation of the pathways. It computes and combines two distances, a "static" distance d_R, taking into account the reactions in

the pathways and a "behavioural" distance d_I, taking into account potential fluxes in the pathways at steady state, as expressed by the T-invariants of the corresponding PNs. Given two pathways represented as PNs, P_1 and P_2, each distance is derived from a corresponding similarity score: $d_X(P_1, P_2) = 1 - score_X(P_1, P_2)$, with $X \in \{R, I\}$.

When computing d_R, the score $score_R(P_1, P_2)$ represents the similarity between the reactions in P_1 and those in P_2. Concretely, each reaction is represented by the enzymes which catalyse it and, in turn, each enzyme is identified by its EC number [40], so that a pathway is reduced to a multiset of EC numbers (a multiset, rather than a set since the same enzyme may have multiple occurrences in a pathway). Then $score_R(P_1, P_2)$ measure the similarity between the multisets of the EC numbers associated to P_1 and P_2, respectively. The similarity between single enzymes is simply the identity of EC numbers, but finer similarity measures could be easily accommodated in this setting. For comparing multisets, the present version of *CoMeta* allows to use either the Sørensen [37] or the Tanimoto [39] indexes extended to multisets.

When computing d_I, the set of minimal T-invariants (Hilbert basis) $\mathcal{B}(P_1)$ and $\mathcal{B}(P_2)$ of the two nets are compared. Each invariant is represented as a multiset of EC numbers, corresponding to the reactions in the invariant, and the similarity between two invariants is given, as before, by their similarity index. The similarity score is computed through a heuristic match between the two Hilbert bases and it represents the similarity of the matching pairs. In *CoMeta* the two distances may be combined: $d_C(P_1, P_2) = \alpha \, d_R(P_1, P_2) + (1 - \alpha) \, d_I(P_1, P_2)$, with $\alpha \in [0, 1]$, to move the focus between reactions and functional components, and two organisms can be compared on n metabolic pathways $P_1, \ldots, P_n$ by considering the average distance over the n pathways. In [21] a detailed explanation of the distances is given and a prototype version of *CoMeta* is presented along with some experiments for validating the proposal. This early version of the tool was running under Windows and Linux and used INA (Integrated Tool Analyser) [38] as an external tool for computing the Hilbert basis. Fewer tuning possibilities were offered, since the prototype modelled the pathways as isolated networks and it used only the Sørensen index in computing the distances.

The current version of *CoMeta* is a user-friendly tool written in Java and running under Linux and Mac. For computing Hilbert bases it resorts to 4ti2 [18], a very efficient tool offered in a software package for algebraic, geometric and combinatorial problems on linear spaces. It resorts to the tool *MPath2PN* for transforming a metabolic pathway into a corresponding PN. The translation is from KGML to PNML [11], a standard format for PNs tools, without ubiquitous substances and enzyme-based, and the user can choose either the isolated or the open PN models of the pathways. *CoMeta* offers a set of integrated functionalities through a graphical user interface as shown

(a) CoMeta main window

(b) Second tab: Generate PNs

(c) Third tab: Compute Distances

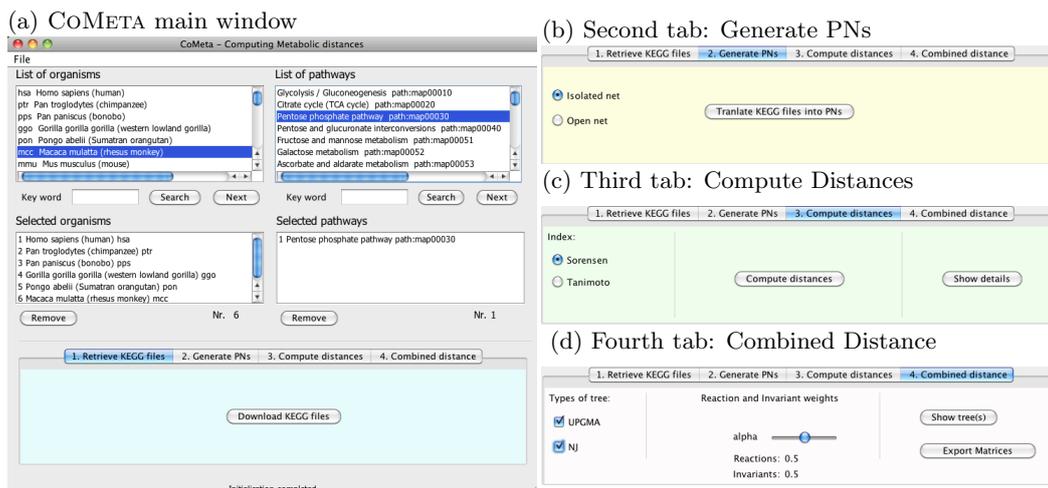

(d) Fourth tab: Combined Distance

Fig. 2. The CoMeta graphical user interface

in Figure 2(a). In the upper part of the window the desired KEGG organisms and pathways can be selected. In the lower part a tabbed panel offers the commands to be performed. The first tab of the panel is shown in the main window, while the others are shown in Figure 2(b), 2(c), and 2(d), respectively. The main functionalities of the tool are the following:

- *Select organisms and pathways* (Figure 2(a), upper part): *CoMeta* proposes the lists of all KEGG organisms and pathways and allows the user to select those to be compared by double-clicking them.

- *Retrieve KEGG information* (Figure 2(a), lower part): *CoMeta* downloads from the KEGG database the selected organisms and pathways.

- *Translate into PNs* (Figure 2(b)): *CoMeta* translates the selected organisms and pathways into corresponding PNs, by using *MPath2PN*. The user can choose to generate either isolated or open PN models of the pathway. CoMeta produces the stoichiometric matrix of the net in a text file which is the input of 4ti2.

- *Compute Distances* (Figure 2(c)): d_R and d_I are computed as described above. The user can select either the Sørensen or the Tanimoto index for multiset comparison. The user can inspect the details of the comparison between any pair of organisms (e.g., T-invariants bases, invariants matches, reactions and invariants scores).

- *Show Phylogenetic trees* (Figure 2(d)): *CoMeta* computes the distance which combines d_R and d_I according to a weight parameter α specified by the user. Such a distance is used to produce and visualise corresponding phylogenetic trees. The user can specify the method to be used for the generation of the phylogenetic trees. Currently *CoMeta* offers the UPGMA [36,35] and Neighbour Joining methods [29,35]. The matrices for d_R, d_I and the com-

bined distance can be exported as text files for further analyses.

4   Conclusions

We presented two tools that we are developing for representing and analysing metabolic pathways through Petri nets. *MPath2PN* is a tool for automatically transforming a metabolic pathway, expressed in one of the various existing databases formats (like KGML or SBML), into a corresponding PN, expressed in one of the formats for PN tools. At present it offers several translations, corresponding to different modelling choices, getting in input a KEGG metabolic pathway and generating a corresponding PN encoded in PNML. *CoMeta* is an interactive tool for comparing metabolic pathways of different organisms. The comparison is based on a distance which may consider both homology of reactions and potential fluxes at steady state.

We are conducting experiments to validate *CoMeta* and our distance. In [21] we already showed that it produces valid phylogenetic classifications and that other measures in the literature, based on more sophisticated representations of a pathway (e.g., using graphs rather than multisets, or considering compounds besides enzymes), do not necessarily give better results than our measure.

This is an ongoing project and we plan to extend both *MPath2PN* and *CoMeta*. We are working on new translations for *MPath2PN*, i.e. from SBML to extended deterministic stochastic PNs (eDSPNs) and coloured stochastic PNs (SCPNs), expressed in the input language of the tool TimeNET [17]. For *CoMeta* we are working in various directions. We are performing extensive explorations of the KEGG database to analyse the significance of our distance. We are experimenting with isolated and open PN models to study the effects of the different modelling choices. Besides, we intend to improve our distance by refining the similarity measures on enzymes, by refining the simple greedy algorithm for matching invariants bases and by associating weights to the pathways, when considering sets of pathways in the comparison.

Acknowledgement

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